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Adsorption and release of thymol using metal organic framework Zn₂(BDC)₂(DABCO) and its chitosan nanocomposite

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8	
 Received: 24 November 2023 Revised: 29 December 2023 Accepted: 4 January 2024 Published online: 10 March 2024 © The Author(s) 2024 © The Author(s) 2024 Metal organic frameworks (MOFs) are new drug delivery systems beca area, tunable pore sizes, and controlled drug release. In the present we MOF is prepared by a quick and simple method. Then, thymol is loaded and its chitosan (MOF-CH) at room temperature. And finally, the results area investigated. Chitosan polymer has been modified the Mathematical activity of the first time and controlled release. The samples were transform infrared (FTIR) spectroscopy for determination of function (XRD) for evaluation of crystal structure, field emission scanning electron investigation of morphology and size, and ultraviolet-visible (UV-Vis) of thymol amount. The antibacterial activity of samples was invest (<i>E. coli</i>) as gram-negative bacterium and <i>Staphylococcus aureus</i> (Staterium. Based on the results, thymol was successfully encapsulated the samples have high-efficiency antibacterial activity. 	ause they have large surface york, Zn ₂ (BDC) ₂ (DABCO) l into MOF (thymol@MOF) elease of thymol from two IOF to develop an efficient re characterized by Fourier al groups, X-ray diffraction on microscope (FESEM) for spectroscopy for evaluation igated by <i>Escherichia coli</i> <i>c. aureus</i>) as gram-positive in MOF and MOF-CH, and

Keywords: Antibacterial activity; Metal organic framework; Thymol; Zn₂(BDC)-2(DABCO) MOF

1. Introduction

Thymol is a monoterpene phenol with antifungal, antioxidant and antibacterial activities [1]. Due to the presence of phenolic hydroxyl group in its structure, thymol is able to destroy the membrane of Gram-negative bacteria and disturb the balance of mineral ions and pH homeostasis inside the cytoplasm of prokaryotic cells. Also, a low concentration of thymol inhibited the release of enterotoxin in Staphylococcus aureus as gram-positive bacterium [2]. The limitation of the use of thymol is due to its low solubility in water and the reduction of its contact with the bacteria in water and, as a result, its inhibition efficiency. Therefore, the expansion of thymol nanocarriers is important [3]. Chitosan is a linear polysaccharide that has various applications due to its unique solubility as well as chemical and biological properties. This compound has many active amino side groups that allow chemical modification and creation of a



Figure 1. Structure of thymol.

wide range of useful derivatives [4]. Chitosan nanoparticles have been widely investigated for drug delivery due to their biodegradability, biocompatibility, and low toxicity [5].

MOFs have recently attracted much attention as a new



Figure 2. FTIR spectrum of (a) MOF, (b) thymol@MOF, (c) MOF-CH, and (d) thymol@MOF-CH.

class of hybrid materials consisting of the self-assembly of metal ions (or clusters) as metal centers and ligands as linkers [6]. These compounds are synthesized by various methods including hydrothermal [7], solvothermal [8], solution [9], ionic liquids [10], son chemical [11], microwave [12], electrochemical [13], diffusion [14], mechanochemical [15], laser ablation [16], combination of ultrasound and microwave [17]. Nowadays, the use of MOF as nanocarriers [18, 19] and compounds with antibacterial activity [20] has been expanded. Recently, there have been reports on thymol loading in MOFs [21, 22]. Zn₂(BDC)₂(DABCO) is a zinc metal-based MOF (Zn-MOF) with nanocarrier applications. This MOF is synthesized by self-assembly of Zn₄O units as metal center and ,4-benzenedicarboxylate and 1,4-diazabicyclo [2.2.2] octane ligands as linker and bridge [23]. Zn₂(BDC)₂(DABCO) MOF can be synthesized by solution and solvothermal method [24, 25]. In the present study, according to functional groups, Zn₂(BDC)₂(DABCO) MOF was investigated as a suitable candidate for thymol absorption. The novelty and objectives of the article are to investigate the possibility of encapsulation and release of thymol as an environmentally friendly volatile antimicrobial



Figure 3. XRD pattern of (a) MOF, (b) thymol@MOF, (c) MOF-CH, and (d) thymol@MOF-CH.



Figure 4. FESEM images of (a) MOF, (b) MOF-CH, and (c) thymol@MOF-CH.

essential oil based on the structure and interactions of pi-pi stacking and hydrogen bond.

2. Experimental procedure

All materials used in this study, including thymol, dimethylformamide (DMF), Zn acetate dehydrate (Zn(OAc)₂.2H₂O), 1,4 benzene dicarboxylic acid, 1, 4 diazabicyclo [2.2.2] octane, and phosphate-buffered saline (PBS), were purchased from Merck Company (Germany), and the distilled water used in this research was the distilled water machine of the Pharmaceutical Sciences Laboratory of Islamic Azad University, Tehran, Iran.

The MOF was prepared by using 0.132 g of Zn acetate dehydrates as the metal center, 0.1 of 1, 4 benzene dicarboxylic acid as the bridging ligand, and 0.035 g of 1, 4 diazabicyclo [2.2.2] octane as the chelating ligand in 25 mL DMF as the solvent under reflux for 30 min at room temperature [24]. The white crystals were rinsed with DMF for removing the remaining ligand and metal and dried under vacuum for 6 hours at 120° C.

To load thymol in the MOF, 5 mL of thymol (100 ppm thymol in the water solution) was added to white MOF crystals (50 mg). The reactants were sealed and stirred at 300 rpm for 30 minutes at room temperature. The solution was centrifuged at 15000 rpm for 10 minutes and dried under vacuum for 6 hours at 60° C. Finally, thymol-MOF was coated with a 2 W% chitosan solution in 2 V% acetic acid, and purified using filtration and dried in a vacuum oven for 6 h at 60° C. According to the structure of thymol (Fig.

1) and MOF [26] and the presence of hydroxyl group and benzene ring in these compounds, hence the interactions of pi-pi stacking and hydrogen bond are the main factors of drug loading according to previous articles [24, 25].

The samples were characterized by FTIR, XRD, FESEM, and UV-Vis spectroscopy. Fourier transform infrared spectroscopy was used to determine the chemical bonds of samples by Spectrum Two FTIR Spectrometer from PerkinElmer. X-ray diffraction pattern was employed for investigation of crystalline structures of samples by of dried suspensions on Si substrate. X-ray diffraction patterns were recorded by STOE X-ray diffractometer with Cu K α radiation ($\lambda = 1.54060$ Å). Morphology and size of samples were evaluated by FESEM (SIGMA VP) microscopes from Zeiss Company. UV-Vis spectroscopy was employed to evaluate the thymol at the maximum wavelength with the Shimadzu model UV-1700 PharmaSpec. The antibacterial activity was evaluated by disk diffusion method against Escherichia coli as gram-negative bacteria (ATCC 25922) and Staphylococcus aureus as gram-positive bacteria (ATCC 25923) by measuring the zone inhibition and minimum inhibitory concentration (MIC).

3. Results and discussion

FTIR spectra of the samples are shown in Fig. 2 in the range of $400 - 4000 \text{ cm}^{-1}$ with KBr pellets. For pure MOF, the absorption peak at $2900 - 3800 \text{ cm}^{-1}$ is due to alkyl C–H, amine O–H, and N–H stretching. In addition, the high and clear intensity peak at 1628 cm^{-1} is assigned to



Figure 5. Percentage of the cumulative thymol release from (a) MOF and (b) MOF-CH.

	Zone inhibition	MIC	Zone inhibition	MIC
Sample	E. coli	S. aureus	E. coli	S. aureus
	(mm)	(mg/mL)	(mm)	(mg/mL)
Thymol	8	3	7	2
MOF	9	1	8	1
Thymol@MOF	11	1	10	1
MOF-CH	21	0.056	20	0.562
Thymol@MOF-CH	25	0.056	23	0.562

Table 1. The zone inhibition and MIC

C=O stretching. At 1390 cm⁻¹, the high intensity peak of C=O belongs to the carboxylic acid groups. It was revealed that the peak at 2370 cm⁻¹ corresponds to environment CO_2 . The C=C stretching of the aromatic bands is observed at 1582 cm⁻¹. The peaks at 1582, 1136, 1100, 819, and 746 cm⁻¹ are assigned with N-C-H compound of DABCO. The results confirm the previous report [24, 25]. For thymol, the peak at 3550 - 3200 cm⁻¹ is assigned with alcohol/phenol O-H stretch. The C-H(aromatic) is assigned at 3150 - 3050 cm⁻¹. The aromatic c=c stretch is assigned at $\sim 1500 \text{ cm}^{-1}$. The CHx deformation band is specified at $1500 - 1400 \text{ cm}^{-1}$. The results confirm the previous report [27, 28]. For chitosan, the strong band at \sim 3300 cm⁻¹ is corresponded to N-H and O-H stretching. The C-H symmetric and asymmetric stretching are observed at around 2921 and 2877 cm⁻¹, respectively. The C=O stretching of amide I is observed at around \sim 1645 $cm^{-1}.$ The C-N stretching of amide III is assigned at 1325 cm^{-1} . The N-H bending of the primary amine is observed at 1589 cm⁻¹. The CH₂ bending and CH₃ symmetrical deformations are confirmed by the presence of bands at around 1423 and 1375 cm⁻¹, respectively. The peak at 1153 cm^{-1} is attributed to asymmetric stretching of the C-O-C bridge. The band at $\sim 1080 \text{ cm}^{-1}$ is corresponded to C-O stretching. The results confirm the previous report [29, 30]. FTIR analysis qualitatively shows the functional groups.

X-ray diffraction patterns of nanostructures in the range of $2\theta = 5^{\circ} - 80^{\circ}$ are presented in Fig. 3. The XRD pattern of the MOF is similar to the previous report and the crystalline structure is preserved after the adsorption based on the previous report [24, 25]. The XRD of chitosan approved the crystalline structure according to the previous report with two characteristic peaks at $2\theta = 10^{\circ}$ and 20° [30]. Therefore, the presence of chitosan reduces the crystal structure. Based on the results, thymol has a crystalline structure [31, 32], and the crystalline structure is reduced due to thymol loading in samples.

FESEM shows shape and size in Fig. 4. Based on these results, the size increases slightly with the increase of thymol and chitosan.

The percentage of the released thymol from MOF and MOF-CH in PBS at 37° C and pH 7.4 [33–35] is shown in Fig. 5. The thymol concentration of the sample was analyzed by UV-Vis spectroscopy. Based on the loading mechanism and release diagram, the drug is loaded in the MOF. According to the results, the presence of chitosan causes a slight increase in the percentage of thymol release. Because chitosan covers the MOF and limits the holes in drug release [36–39]. Therefore, the controlled release system was obtained by increasing the release time of thymol in the presence of chitosan.

The zone inhibition and MIC are shown in Table 1. The medium was used overnight for the growth of *Staphylococcus aureus* and *Escherichia coli* at 37° C for 24 h. Based on the results, the presence of chitosan and thymol increased the antibacterial activity. The results confirm the previous reports [40–42].

4. Conclusion

In this study, thymol was entrapped in the $Zn_2(BDC)_2(DABCO)$ and coated with CH for DDS. Also, these samples showed good release performance of thymol in PBS at pH 7.4. Its structural properties were characterized by FTIR, XRD, and FESEM. The release of thymol from the samples was confirmed by UV-Vis spectroscopy. The samples showed good antibacterial activity against *E. coli* as gram-negative bacteria and *S. aureus* as gram-positive bacteria. Based on the results, the presence of chitosan and thymol has a great effect in increasing antibacterial activity. Therefore, the future perspective based on these compounds can have a good potential for the development of their antibacterial application.

Ethical approval

This manuscript does not report on or involve the use of any animal or human data or tissue. So the ethical approval is not applicable.

Authors Contributions

All the authors have participated sufficiently in the intellectual content, conception and design of this work or the analysis and interpretation of the data (when applicable), as well as the writing of the manuscript.

Availability of data and materials

Data presented in the manuscript are available via request.

Conflict of Interests

The author declare that they have no known com-

peting financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Motakef Kazemi et al.

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