

Contents available at ISC and SID

Journal homepage: www.rangeland.ir



Full Length Article:

Effects of Selection on Genetic Parameters of Secale montanum Based on Seed Storage Protein Marker

Parvin Salehi Shanjani^A, Ali Ashraf Jafari^B, Roya Hoseinzadeh^C

Received on: 30/08/2013 Accepted on: 29/10/2013

Abstract. Secale montanum is one of the important perennial grasses growing naturally in arid to semiarid pastures and rangelands, with a typical Mediterranean climate, in northern and western Iran at altitudes of 800-2900 m. In this paper, seed storage protein profiles of nine wild populations of S. montanum from different regions of Iran and their phenotypically superior progenies as well as a multi-origin polycross (PLC) were studied. High levels of polymorphism were observed over all populations with the average number of bands and average heterozygosity. Superior progeny of different populations showed less genetic variability than wild parents in terms of band diversity, whereas PLC samples showed extremely high values of genetic parameters. Two locally common bands were observed in almost all wild parent populations, which are missing in superior progeny of different populations and PLC. These results provide highly support for the hypothesis that neutral genetic diversity has been reduced or inadvertently lost via artificial selection. Among wild parent populations and their superior progenies significant differences were observed in expected heterozygosity suggesting that more intensive breeding practices may have resulted in a further erosion of genetic variability. Neighbor-joining cluster analysis showed that wild populations and the phenotypically superior progeny of different populations were separated into two groups. This suggests that founder effects and subsequent selection have had more effect on the genetic differentiation between these accessions than geographical separation. This technology, seed storage protein profiling, has great potential for use in breeding programmes.

Key words: *Secale montanum*, Seed storage proteins, Superior progeny, Wild population

^AAssistant Professor, Natural Resources Gene Bank, Research Institutes of Forests and Rangelands, Tehran, Iran. (Corresponding Author). E-mail: psalehi1@gmail.com

^BProfessor, Natural Resources Gene Bank, Research Institutes of Forests and Rangelands, Tehran, Iran. ^CAzad University of Karadj, Karadj, Iran.

1. Introduction

Secale montanum Guss. (syn. S. dalmaticum Vis., common names: secale mountain rye) is outbreeding, perennial grass, widely distributed in central and southern Europe, northern Africa, Asia Minor, Transcaucasia, Iraq, Iran, and northern Pakistan (Davis, 1985). It normally inhabits dry, stony, or rocky hillsides, and as a weed in crops of Triticum turgidum Steud (Davis, 1985). S. montanum is one of the important perennial grasses naturally growing in arid to semiarid pastures and rangelands, with a typical Mediterranean climate, in northern and western Iran at altitudes of 800-2900 m. It is used for grazing and hav production as well as revegetating the overgrazed semi-steppe rangelands (Peymani-Fard, 1993). Because of its dense network of roots, S. montanum is recommended as a part of a seed mix for erosion control (Anderson and Brooks, 1975; Mclean and Clark, 1980). A few studies have been conducted on S. montanum in different ecological conditions of Iran and revealed that there was considerable variation in herbage yield, seed yield and crude protein content (Rahmani et al., 2002). Although S. montanum is described as a short-lived, palatable, leafy, perennial which can provide winter grazing in subtropical areas with fair winter rainfall (Rahmani et al., 2002), it has some troublesome characteristics including small seed size, shattering and pre-harvest sprouting (Gazanchian, 2006). Gordon-Werner and Dorffling (1988) have studied some of the mechanisms of drought tolerance in mountain rye. It is also used as a source of disease resistance genes (Andriyash, 1989) and cytoplasmic male sterility (Lapinski, 1991) in cereal rye breeding programmes in Eastern Europe. Attempts to breed a perennial grain type from hybrids between S. cereale and S. montanum were initially hindered by interchanges among three pairs

chromosomes, but eventually were successful (Reimann-Philipp, 1995).

The most frequent breeding methods applied to crop species involve different forms of mass selection, recurrent phenotypic selection development of synthetic populations. Information about germplasm diversity and relationships among elite breeding materials is of fundamental importance in plant breeding (Hallauer and Miranda, 1988). This is especially true for species like S. montanum which suffers severe depression (Geiger inbreeding Miedaner, 1999). However, there is neither information on the genetic quality of wild S. montanum populations nor information on the progeny to be used in breeding programmes. Reports of studies based on different plant species provide conflicting results on the impact of domestication on the genetic diversity of populations (Chaisurisri and El Kassaby, 1994; Rajora, 1999; Moran et al., 2000; Godt et al., 2001; Icgen et al., 2006). Also the impact of domestication on the genetic diversity of progeny populations is poorly understood (Stoehr and El-Kassaby, 1997; Schmitdtling and Hiplins, 1998). Such studies on genetic diversity of initial selection materials are essential for successful breeding and creation of new cultivars.

S. montanum has been studied by morphological (Fredriksen and Petersen, 1998; Rahmani et al., 2002; Oram, 2013), cytological (Katsumasa et al., 1990; Petersen and Doebley, 1993, Petersen et al., 2004; Cuadrado and Jouve, 1997, 2002; Riley, 1955; Sheidai, 2008) isozymes (Vence et al., 1987a, b), restriction fragment length polymorphisms of plastid genome (cpDNA RFLPs) (Murai et al., 1989), restriction fragment length polymorphisms of mitochondrial DNA (mDNA RFLPs) (Skizai et al., 2007), rDNA spacer-lengths (Reddy et al., 1990), ITS of the 18 s-5.8 s rDNA (De Bustos and Jove, 2002), Random Amplified Polymorphic (RAPDs) (Del Pozo *et al.*, 1995), Amplified Fragment Length Polymorphisms (AFLPs) (Chikmawati *et al.*, 2005), ISSR and SCAR markers (Vaillancourt *et al.*, 2008) and microsatellite (Shang *et al.*, 2006; Jenabi *et al.*, 2011) analyses. The genetic structure of the Iranian *S. strictum* populations, however, still remains unclear despite its usefulness as a genetic resource.

Characterization of germplasm using biochemical fingerprinting has got special attention. The protein profiling of germplasm and use of genetic markers have been widely and effectively used to determine the taxonomic and evolutionary aspects of several crops (Murphy et al., 1990; Khan, 1990; Das and Mukarjee, 1995; Ghafoor et al., Sodium Dodecyl 2002). Sulphate Polyacrylamide Gel Electrophoresis (SDS-PAGE) is most economical, simple extensively used biochemical technique for analysis of genetic structure of germplasm. Leaf total proteins have been used as genetic markers in breeding programs (Hirano, 1982; Vries, 1996; Kamel and Hassan, 2001; Reddy and Munirajappa, 2005; Mohamed et al., 2006), as well as in basic studies on population genetics (Torkpo et al., 2006) and reproductive biology (Cardeña et al., 1998). To our knowledge, no studies have yet been made in Iran on the diversity of S. montanum germplasm based on total protein electrophoresis. We therefore address two issues in the present study: (i) the structure of genetic diversity in different wild S. montanum populations, and (ii) the impact of selection on genetic diversity of progeny populations using seed storage proteins.

2. Materials and Methods

Seed material of nine wild populations of *Secale montanum* (accession No.: 467, 1275, 2283, 2292, 1549, 591, 2382, 1567 and 941) and their phenotypically superior progenies as well as the multi-

origin polycross (PLC), provided from Iranian Natural Resources Gene Bank (INRGB), were used in the present study (a total of 190 entries).

For study of the extent of genetic variation based on SDS-PAGE markers, a total of 190 entries were selected from nine wild populations, their superior progenies and PLC (10 plants for each population). Preliminary experiments (data not shown) indicated that a larger sample (20 plants for each population) did not modify the results substantially regarding the amount or the structure of polymorphism. Seed storage proteins were extracted using 0.05M Tris-HCL pH=8, 0.2% SDS, 5M urea, 1% Bmercaptoethano 1.

Electrophoresis was carried out in discontinuous Sodium the polyacrylamide dodecylsulphate gel electrophoresis (SDS-PAGE) system of Laemmli (1970) using 12% separating gel and 5% (w/v) stacking gel (Fig. 1). The molecular weights of the dissociated protein were estimated by using molecular weight standard proteins "MW-SDS-70 Kit". Gels were gently shaken until the background of the gel became clear and polypeptide bands were clearly visible.

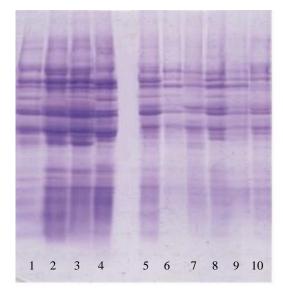


Fig. 1. Seed storage protein band profiles of the multi-origin polycross (PLC) samples (1-4) and the superior progenies (5-10) of S. montanum

2.1. Data analysis

For protein profile data, to avoid taxonomic weighting, the intensity of bands was not taken into consideration, only the presence of bands was taken as indicative. The scores were 1 for the presence and 0 for the absence of a band. Then, the indices of genetic diversity, such as the number of bands (Na), locally common bands with frequency $\leq 25\%$ (Nalc), Percentage of Polymorphic Loci (PPL) and expected heterozigosity (He), were calculated using POPGENE 32 software (Yeh et al., 1999) on the basis of gene frequencies. At the same time, the genetic structure within and among populations were detected using the software AMOVA- PREP 1.01 (Miller, 1997) and WINAMOVA (Excoffier, 1995) in order to partition the genetic variation among local and exotic groups, among populations within groups and among individuals within populations. The significance of each variance component was tested with permutation tests (Excoffier et al., 1992). Genetic distances were estimated according to Nei (1978) and the resulting similarity matrix was subjected to Principal Component Analysis (PCA), UPGMA algorithm using NTSYS-pc 2.01 (Rohlf, 2004), and Neighbor- Joining (NJ) analysis using MEGA4 software (Tamura et al., 2007). Wright's Fst and Nm were used to estimate population differentiation and gene flow, respectively. The rate of gene flow was estimated indirectly from the proportion of total diversity that was found among populations (Wright, 1931, 1951). A 999 random permutation Mantel test (Gower 1966) was used to assess the correlation between the calculated distance matrices.

3. Results

On the basis of the relative mobility of on the proteins gel, polypeptide bands of different sizes ranging from 6.606 to 269.153 kDa, from nine wild parent populations, their superior progenies and PLC samples of montanum, S. identified. Different populations showed quite different band frequency distributions among wild parent. progeny and PLC genotypes (Fig. 2). Among the nine wild populations, the mean PPL and He values were 62.50% and 0.253, respectively. Population p-Bojnurd (Na, PPL and He values: 31, 81.25% and 0.345, respectively) and p-Zanjan1 (Na, PPL and He values: 32, 87.50% and 0.332, respectively) had the highest level of variability, whereas population p-Zanjan3 had the lowest level of variability (Na, PPL and He 18.75% values: 27, and 0.079, respectively) (Table 1). Among the superior progenies, the mean PPL and He values were 56.45% and 0.221, respectively. Within the superior progenies, PPL value ranged from 37.50% (s-Karaj3) to 71.88% (s-Karaj1), He value ranged from 0.116 (s-Esfahan) to 0.71.88 (s-Zanjan) (Table 1). The PPL and He values of PLC (93.75% and 0.409, respectively) were higher than all the wild parent and superior populations. Almost in all wild populations (except p-Zanjan3) two locally common bands (with frequency $\leq 25\%$) were observed, superior missing in progeny different populations and PLC. Comparison of different wild populations with their superior progenies showed significant decrease in the genetic parameters of superior progenies in the most populations.

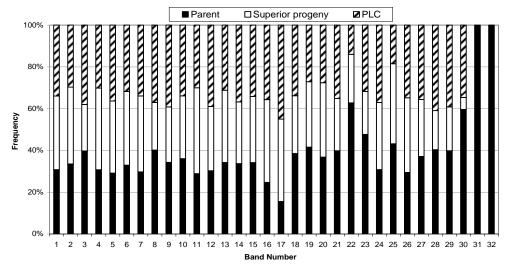


Fig. 2. Seed storage protein band frequencies of the wild parent, the superior progenies and multi-origin polycross (PLC) samples of *S. montanum*

Table 1. Genetic diversity parameters of the wild parent populations, the superior progenies of

different populations and PLC of S. montanum

	Na		Nalc		PPL		Не		
	Parent	Superior Progeny	Parent	Superior Progeny	Parent	Superior Progeny	Parent	Superior Progeny	
467	32	25	2	0	87.50	68.75	0.332	0.311	
1275	32	29	2	0	59.38	68.75	0.256	0.274	
2283	27	25	0	0	18.75	50.00	0.079	0.217	
2292	32	29	2	0	71.88	68.75	0.319	0.285	
1549	31	26	2	0	81.25	46.88	0.345	0.190	
591	31	27	2	0	62.50	37.50	0.250	0.163	
2382	31	29	2	0	78.13	71.88	0.293	0.265	
1567	32	22	2	0	46.88	43.75	0.181	0.116	
941	32	27	2	0	56.25	50.00	0.222	0.166	
Mean					62.50	56.45	0.253	0.221	
PLC		30		0		93.75%		0.409	

Na = observed number of bands; Nalc = Number of locally Common Bands (frequency <= 50%); He = Nei's gene diversity; PPL = Percentage of polymorphic loci

The coefficient of genetic differentiation (Fst) among wild parent populations was 0.3 (31% of total genetic variation resided among, and 69% within populations). Among superior the progenies of different populations, Fst was 0.360. The level of gene flow (Nm) between wild parent populations was 1.373. Fst between wild parent populations and their superior progenies was 0.363; indicating only about 20% genetic variation resided among the three groups (wild parents, superior progenies and PLC). The level of gene flow (Nm) between superior progenies and their wild

progenitors was 0.714 individuals per generation. AMOVA analysis showed that the variation among the wild parent populations and within the populations accounted for 31% and 69% of the total variation, respectively (P < 0.01) (Table 2). In the progeny data set, these values were 38% and 62% (P<0.01),**AMOVA** respectively. analysis revealed a highly significant genetic differentiation (P < 0.01) among the wild parent populations and their progenies. Of the total genetic diversity, 20% was attributable to between-group diversity and the rest (80%) to differences within groups (Table 2).

Table 2. Analysis of Molecular Variance (AMOVA) of the wild parent populations, the super	ior
progenies of different populations and PLC of S. montanum based on total protein profile	

Source	Degrees of Freedom	Sum of Squares	Mean of Squares	Est. Var.	Variation %	P
Among groups (parents, superior progenies and PLC)	2	158.354	79.177	1.412	20%	
Within groups	187	1044.567	5.586	5.586	80%	0.001
Among parent populations	8	180.578	22.572	1.851	31%	
Within parent populations	81	329.000	4.062	4.062	69%	0.010
Among superior progeny populations	8	199.289	24.911	2.140	38%	
Within superior progeny populations	81	284.100	3.507	3.507	62%	0.010

The genetic distance of the populations using Nei's genetic distances is shown in (Table 3). The data ranged from 0.041 (P-1275 and P-2292) to 0.703 (P-1567 and S-1567), with an average of 0.279 (Table 3). To elucidate the genetic relationships among wild parent populations, superior progeny populations and PLC samples, an UPGMA dendrogram was produced using Nei's genetic distances (Fig. 3). The nine parent populations and their superior populations were grouped into three clusters, cluster I having populations including PLC, the superior progenies of almost all populations and two wild parent populations, cluster II consisted of wild parent populations, and cluster III, having only one population (S-1567).

The total protein data were also used for conducting Principal Coordinate Analysis (PCoA) to study further the genetic diversity among the nine wild parent *S. montanum* populations and their progenies (Fig. 4). The results of the PCA showed that most of the wild populations were clearly separated from their progenies (Fig. 4). The first three principal coordinates accounted for 80% of the total variation among the populations or progenies.

Correlation coefficients among pairwise genetic distance matrices of wild parent samples and their superior progenies were calculated using mantel's test. Regression and correlation analysis between genetic distances showed very low correlation, but significant (p < 0.04) (Fig. 5).

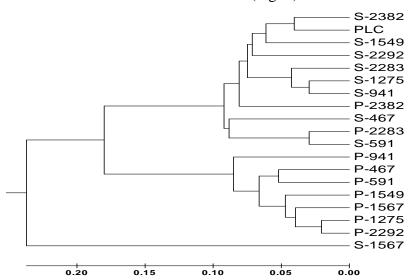


Fig. 3. Phenograms of the wild parent populations (with P prefix), the superior progenies of different populations (with S prefix) and multi-origin polycross (PLC) of S. montanum based on seed storage protein profiles

Table 3. Pairwise values for Nei's genetic distances of the wild parent populations (with P prefix), the superior progenies of different populations (with S prefix) and PLC of S. montanum

	P-467	P-1275	P-2283	P-2292	P-1549	P-591	P-2382	P-1567	P-941	S-467	S-1275	S-2283	S-2292	S-1549	S-591	S-2382	S-1567	S-941
P-467	0.000																	
P-1275	0.175	0.000																
P-2283	0.398	0.334	0.000															
P-2292	0.126	0.041	0.302	0.000														
P-1549	0.096	0.077	0.364	0.097	0.000													
P-591	0.104	0.087	0.370	0.074	0.104	0.000												
P-2382	0.275	0.287	0.213	0.257	0.251	0.280	0.000											
P-1567	0.224	0.066	0.329	0.092	0.108	0.173	0.320	0.000										
P-941	0.186	0.156	0.321	0.153	0.142	0.186	0.266	0.197	0.000									
S-467	0.197	0.073	0.307	0.101	0.126	0.157	0.341	0.122	0.143	0.000								
S-1275	0.179	0.108	0.305	0.104	0.139	0.157	0.268	0.133	0.144	0.089	0.000							
S-2283	0.318	0.334	0.405	0.388	0.316	0.392	0.347	0.343	0.274	0.219	0.239	0.000						
S-2292	0.331	0.210	0.277	0.243	0.253	0.306	0.303	0.242	0.198	0.137	0.076	0.125	0.000					
S-1549	0.314	0.133	0.203	0.118	0.236	0.227	0.288	0.131	0.156	0.156	0.115	0.383	0.162	0.000				
S-591	0.240	0.122	0.279	0.136	0.204	0.192	0.352	0.146	0.197	0.043	0.098	0.193	0.135	0.157	0.000			
S-2382	0.373	0.260	0.353	0.276	0.339	0.356	0.284	0.331	0.264	0.171	0.127	0.195	0.077	0.197	0.190	0.000		
S-1567	0.224	0.163	0.275	0.168	0.201	0.230	0.295	0.189	0.117	0.122	0.130	0.173	0.118	0.118	0.129	0.132	0.000	
S-941	0.287	0.239	0.419	0.227	0.304	0.272	0.300	0.303	0.232	0.211	0.097	0.252	0.141	0.293	0.159	0.212	0.251	0.000
PLC	0.222	0.246	0.148	0.243	0.217	0.254	0.095	0.282	0.224	0.204	0.078	0.130	0.100	0.091	0.179	0.081	0.361	0.142

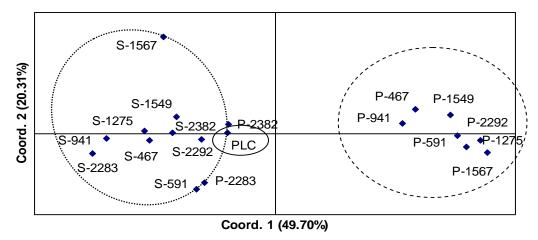


Fig. 4. Two-dimensional graph based on the ordination scores of the principal coordinate analysis of the wild parent populations (with P prefix) and the superior progenies of different populations (with S prefix) and PLC of S. montanum

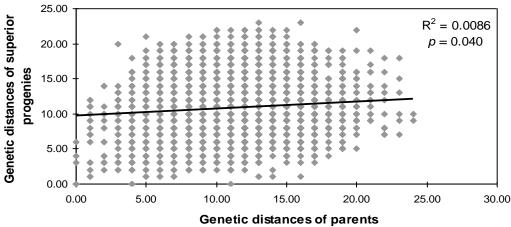


Fig. 5. Correlations between genetic distances among the wild parent populations and the superior progenies of different populations of *S. montanum*

4. Discussion

Loss of genetic diversity and increased population differentiation from source populations common are problems associated with breeding programmes established from a small number of founders. Like many other studied plant species where cultivars have lower genetic diversity than their wild relatives (Doebley, 1989; Gepts, 2004; Zhou et al., 2005; Miller and Schaal, 2006; Wu et al., 2006; Salehi Shanjani et al., 2013), the progenies superior of different montanum populations maintain lower levels of genetic diversity as their parents (mean He value for wild parent populations was significantly higher than that of their progenies). The heterozygosity of PLC samples was

higher compared to all wild parent or superior progeny populations. Despite the retention of genetic diversity in PLC, a detectable shift in gene frequency was revealed by the distribution of band frequencies. These results demonstrates that artificial breeding practices result in a decrease in genetic variability in terms of band diversity but which is not necessarily detectable from levels of heterozygosity.

Selective breeding often produces an improvement in phenotype. Artificial selection can separate adult individuals from a parent generation into two groups, those selected and those to be discarded, based on the characteristics that are determined by the changes in the gene frequency (Li *et al.*, 2010). This has been

confirmed in many species, such as Bluebunch Wheatgrass (Larson et al., 2000), maze (Labate et al., 1997; Zou et al., 2010), Cassava (Oslen and Schall, 1999; Manu-Aduening et al., 2013) and rice (Virk et al., 2003). In the present study, significant genetic differentiation among the wild parent populations and their superior progenies is also due to band frequency alterations. The most striking change in band frequencies is the loss of low frequency bands, which is proved to be a common phenomenon in cultivars as a consequence of small population size, genetic drift, selection (Gosling, 1982; Dillon and Manzi, 1987; Launey et al., 2001; Zhang et al., 2004). The major reason for the genetic differentiation between the wild parent populations and their progenies in this study appears to be artificial selection as the superior progenies have extensively selected. been investigation further demonstrates that gene frequency change is the genetic basis of character improvement selective breeding.

diversity Genetic is always changing, but the report on the state of the worlds plant genetic resources (FAO, 1996), summarizing country reports, suggests that "recent losses of diversity have been large, and that the process of erosion continues." It points out that while loss of genes is of particular concern, loss of gene complexes and unique combinations of genes (as in different landraces) can also have important consequences. Genetic erosion may thus be defined as a permanent reduction in richness or evenness of common localized alleles or the loss of combination of alleles over time in a defined area. This definition recognizes diversity has two distinct components in (i) the number of different entities and (ii) their relative frequencies. It also suggests that it is specifically loss of locally adapted alleles that is most significant. Two locally common bands

were detected in almost all wild parent populations, which is very important part of genetic diversity, are missing in superior progenies and PLC. This process considered as genetic diversity erosion. Genetic erosion will be detrimental to the short-term viability of individuals and populations, the evolutionary potential of populations and species, and the direct use of genetic resources (Brown et al., 1997). Recent genetic erosion and/or the risk of imminent genetic erosion are key factors in determining the priority given different areas for conservation interventions whether ex situ, in situ or a combination of both.

conclusion, In this study demonstrates the high levels of polymorphism detectable with seed storage proteins even within superior progenies of S. montanum. technology has the potential to be of great use in monitoring levels of genetic variation within wild populations as well for parentage and relatedness as Between wild purposes. parent populations and their progenies significant differences were observed in expected heterozygosity suggesting that more intensive breeding practices may have resulted in a further erosion of genetic variability. These results also show that allelic diversity is a more sensitive measure of differences in genetic variation between wild and populations than progeny overall heterozygosity. This is likely to be a result of the loss of low frequency alleles when new populations are created from larger founder ones. These results provide highly support for the hypothesis that neutral genetic diversity has been reduced or inadvertently lost via artificial selection. Neighbor-joining analysis showed that wild populations and the phenotypically superior progeny of different populations were separated into two groups. This suggests that founder effects and subsequent selection have had more effect on the genetic

differentiation between these accessions than geographical separation. Differences in genetic variation observed among superior progenies may be a result of geographical separation of their parent populations. This technology has great potential for use in breeding programmes.

5. Acknowledgement

This work was supported by the Research Institute of Forests and Rangelands (RIFR), Iran; Project no. 12-09-09-7901-87001.

Literature

- Anderson, E. W., and Brooks, L. E., 1975. Reducing erosion hazard on a burned forest in Oregon by seeding. *Jour. Range Manage*. **28**(5): 394-398.
- Andriyash, V. P., 1989. Breeding winter rye and the differentiation of varieties for resistance to root rots in the Poles'e area of the Ukrainian SSR, Russian. English summary in Plant breeding abstracts. **60:** 912-919.
- Brown, A., Young, A., Burdon, J., Christides, L., Clarke, G., Coates, D., and Sherwin, W., 1997. Genetic indicators for state of the environment reporting., Australia State of the Environment Technical Paper Series (Environmental Indicators), Department of Environment, Sport and Territories, Canberra.
- Cardeña, R., Oropeza, C., and Zizumbo, D., 1998. Leaf proteins as markers useful in the genetic improvement of coconut palms, Euphytica. **102**: 81–86.
- Chaisurisri, K., and El Kassaby, Y. A., 1994. Genetic diversity in seed production population vs natural populations of *Picea sitchensis*. Biodiversity Conserv. **3:** 512–523.
- Chikmawati, T., Skovmand, B., and Gustafson, J. P., 2005. Phylogenetic relationships among *Secale* species revealed by amplified fragment length polymorphisms. Genome, **48:** 792-801.
- Cuadrado, A., and Jouve, N., 1997. Distribution of highly repeated DNA sequences in species of the genus *Secale*. Genome. **40:** 309-317.
- Cuadrado, A., and Jouve, N., 2002. Evolutionary trends of different repetitive DNA sequences during speciation in the genus *Secale. Jour. Hered.* **93:** 339-345.
- Das, S., and Mukharjee, K. K., 1995. Comparative study on seed proteins of Ipomoea. Seed Sci. Tech. **23:** 501–509.

- Davis, P. H., 1985. Flora of Turkey and the eastern Aegean Islands, Vol. 9. Edinburgh, The University Press. Pp. 256-257.
- De Bustos, A., and Jouve, N., 2002. Phylogenetic relationshios of the genus *Secale* based on the characterization of rDNA ITS sequences. Pl Syst. Evol. **235**: 147-154.
- Del Pozo, J. C., Figueiras, A. M., Benito, C., and De La Pena, A., 1995. PCR derived molecular markers and phylogenetic relationships in the *Secale* genus. Biologia Plant. **37:** 481-489.
- Dillon, R. T., and Manzi, J. J., 1987. Hard clam, *Mercenaria mercenaria*, brood stocks: genetic drift and loss of rare alleles without reduction in heterozygosity. Aquaculture, **60**: 99–105.
- Doebley, J., 1989. Isozymic evidence and evolution of crop plants. In: Soltis, E. D., Soltis, P. M., eds, Isozymes in Plant Biology, Dioscorides, Portland, Oregon, 165–191.
- Excoffier, L., 1995. AMOVA 1.55 (Analysis of Molecular Variance). University of Geneva, Switzerland, Genetics and Biometry Laboratory.
- Excoffier, L., Smouse, P., and Quattro, J., 1992. Analysis of molecular Variances among DNA restriction data. Genetics, **131:** 479-491.
- FAO, 1996. Report on the state of the world's plant genetic resources for food and agriculture. FAO, Rome, 75 pp.
- Frederiksen, S., and Peterson, G., 1998. A taxonomy revision of *Secale* (Triticeae, Poaceae). *Nordic Jour. Bot.* **18:** 399-420.
- Geiger, H. H., and Miedaner, T., 1999. Hybrid rye and Heterosis. In Genetics and exploitation of Heterosis in crops. Edited by Coors JG, Pandey S. Madison, Wisconsin: Crop Sci. Soc. America; **1999**: 439–450.
- Gepts, P., 2004. Crop domestication as a long-term selection experiment. Plant Breed. Rev. **24:** 1–44.
- Ghafoor, A., Ahmad, Z., Qureshi, A. S., and Bashir, M., 2002. Genetic relationship in *Vigna mungo* (L.) Hepper and *V. radiata* (L.) R. Wilczek based on morphological traits and SDS PAGE. Euphytica, **123**: 367–378.
- Ghazanchian, A., 2006. Factors affecting seed shattering in *Secale montanum* and improvement of management in seed production sites. Final Technical Report of Research Institute of Forests and Rangelands No. 6327, 54 p. (In Persian).
- Godt, M. J., Hamrick, J. L., Edwards-Burke, M. A., and Williams, J. H., 2001. Comparison of genetic diversity in white spruce (*Picea glauca*)

- and jack pine (*Pinus banksiana*) seed orchards with natural populations. Can. *Jour. For. Res.* **31:** 943–949.
- Gordon-Werner, G., and Dorffling, K., 1988. Osmotic adjustment and stomatal characteristics of the *Secale cereale x Secale montanum* cross Permontra. *Jour. Agron. crop sci.* **161:** 30-39.
- Gosling, E. M., 1982. Genetic variability in hatchery-produced Pacific oyster (*Crassostrea gigas* Thunberg). Aquaculture, **26:** 273–287.
- Gower, J. C., 1966. Some distance properties of latent root and vector methods used in multivariate analysis. Biometrika, **53**: 325-338.
- Hallauer, A. R., and Miranda, J. B., 1988. Quantitative genetics in maize breeding. 2nd ed. Iowa State University Press. Ames, IA.
- Hirano, H., 1982. Varietal differences of leaf protein profiles in mulberry. Phytochemistry. **21:** 1513–1518.
- Icgen, Y., Kaya, Z., Cengel, B., Velioglua, E., Ozturka, H., and Onde, S., 2006. Potential impact of forest management and tree improvement on genetic diversity of Turkish red pine (*Pinus brutia* Ten.) plantations in Turkey. For. Ecol. Manage. **225**: 328–336.
- Jenabi, T., Sajedi, H., and Rahiminejad, M. R., 2011. Biodiversity of *Secale strictum* in Iran measured using microsatelites. Genet. Resour. Crop. Evol. **58:** 497-505.
- Kamel, E. A., and Hassan, L. M. A., 2001. The significance of cuticular features, petiole anatomy and SDS-PAGE in the Taxonomy of the Lauraceae. *Pakistan Jour. Biol. Sci.* **4:** 1094–1100.
- Katsumasa, N., Shoji, O., and Sadao, S., 1990. B chromosomes of *Secale cereale* L. and *S. montanun* G. Ikushugaku zasshi, **40:** 147-152.
- Khan, M. K., 1990. Production and utility of chickpea (*Cicer arietinum* L.) in Pakistan. Progressive Farming. **10:** 28-33.
- Labate, J. A., Lamkey, K. R., Lee, M., and Woodman, W. L., 1997. Molecular Genetic Diversity after Reciprocal Recurrent Selection in BSSS and BSCB1 Maize Populations. Crop Sci. 37: 416-423.
- Laemmli, U. K., 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. Nature, **227**: 680-685.
- Lapinski, M., 1991. Breeding and hybrid studies of the cytoplasmic pollen sterility system in rye. Polish, English abstract in Plant breeding abstracts, **63**: 1073-1074.

- Larson, S. R., Jones, T. A., Hu, Z. M., McCracken, C. L., and Palazzo, A., 2000. Genetic Diversity of Bluebunch Wheatgrass Cultivars and a Multiple-Origin Polycross. Crop Sci. **40**: 1142–1147.
- Launey, S., Barre, M., Gerard, A., and Naciri-Graven, Y., 2001. Population bottleneck and effective size in Bonamia ostreae-resistant populations of *Ostrea edulis* as inferred by microsatellite markers. Genet. Res. **78**: 259–270.
- Li, H. D., Liang, Y. Z., Xu, Q. S., and Cao, D. S., 2010. Model population analysis for variable selection. *Jour. Chemometrics*, **24**: 418–423.
- Manu-Aduening, J. A., Peprah, P. P., and Agyeman, A., 2013. Genetic variability of cassava Progenies developed through introgression of cassava mosaic disease resistance into ghanaian landraces. *Jour. Crop Sci. Biotech.* **16:** 23-28.
- McLean, A., and Clark, M. B., 1980. Grass, trees, and cattle on clearcut-logged areas. *Jour. Range Manage.* **33(3):** 213-217.
- Miller, A. J., and Schaal, B. A., 2006. Domestication and the distribution of genetic variation in wild and cultivated populations of the Mesoamerican fruit tree *Spondias purpurea* L. (Anacardiaceae). Mol. Ecol. **15:** 1467–1480.
- Miller, M. P., 1997. AMOVA-PREP, a program for the preparation of AMOVA input files for use with WINAMOVA. Department of Biological Sciences, Northern Arizona University, Flagstaff, AZ.
- Mohamed, T. R., S. F. Khalifa, and R. M. Salah EL-Dine. 2006. Leaf Protein Electrophoretic Profiles and Chromosome Numbers of Some Araceae. *Int. Jour. Agri. Biol.* **8:** 231–234.
- Moran, G. F., Butcher, P. A., and Glaubitz, J. C., 2000. Application of genetic markers in domestication conservation and utilization of genetic resources of Australasian tree species. *Aust. Jour. Bot.* **48:** 313–320.
- Murai, K., Naiyu, X. U., and Tsunewaki. K., 1989. Studies on the origin of crop species by restriction endonuclease analysis of organellar DNA: III. Chloroplast DNA and interspecific relationships in the genus *Secale. Japanese Jour. Genet.* **64:** 35–47.
- Murphy, R. W., Sites, J. W., Buth, D. G., and Haufler, C. H., 1990. Protein I: Isozyme Electrophoresis., In: Hillis, D. H., and Moritz, C., eds., Molecular Systematics, Sinnauer Association, Sunderland, Massachusetts, Pp. 45-126.

- Nei, M., 1978. Estimation of average heterozygosity and genetic distance from a small number of individuals. Genetics, **89:** 583-590.
- Oram, R. N., 2013. *Secale montanum -* a wider role in Australasia?, New Zealand J. Agri. Res. 39: 629-633.
- Oslen, K. M., and B. A. Schaal. 1999. Evidence on the origin of cassava: phylogeography of *Manihot esculenta*. Proc. Natl. Acad. Sci. USA **96:** 5586-5591.
- Petersen, G., and Doebley, J. F., 19930. Chloroplast DNA variation in the genus *Secale* (Poaceae). Plant Syst. Evol. **87:** 115–125.
- Petersen, G., Seberg, O., Aagesen, L., and Frederiksen, S., 2004. An empirical test of the treatment of indels during optimization alignement based on the phylogeny of the genus *Secale* (Poaceae). Mol. Phylogenet. Evol. **30**: 733–742.
- Peymani-Fard, B., 1993. A study on the promising ecotypes of *Secale montanum* Guss. Proceedings of the XVII International Grasssland Congress, P: 199-200.
- Rahmani, E., Jafari, A. A., and Hedaiati, P., 2002. Evaluation of seed yield, forage yield and their components in Mountain Rye (*Secale Montanum* Guss.) through correlation, regression and path analysis. Iranian Mol. Phylogenetics Evol. **30:** 733–742. *Jour. Rangelands For. Plant Breeding Genet. Res.* **10:** 2-12. (In Persian).
- Rajora, O. P., 1999. Genetic biodiversity impacts of silvicultural practices and phenotypic selection in white spruce. Theor. Appl. Genet. **99:** 954–961.
- Reddy, P., Appels, R., and Baum, B. R., 1990. Ribosomal DNA spacer-length variation in *Secale* spp. (Poaceae). Plant Syst. Evol. **171**: 203-220.
- Reddy, P. M., and Munirajappa, M., 2005. Electrophoretic studies in induced mutants of diploid mulberry genotype S13. *Indian Jour. Biotech.* **4:** 422-423.
- Reimann-Philipp, R., 1995. Breeding perennial rye. Plant breeding rev. **13:** 265-292.
- Riley, R., 1955. The cytogenetics of the differences between some *Secale* species. *Jour. Agri. Sci.* **46:** 377–383.
- Rohlf, J. F., 2004. NTSYS-pc: Numerical taxonomy and multivariate analysis system, version 2.11., Exeter, Setauket, NY.
- Salehi Shanjani, P., Jafari, A. A., and Calagari, M., 2013. Genetic variation among wild and

- cultivated *Agropyron desertorum* populations based on total protein profiles and phenotypic traits. *New Zealand Jour. Crop Horti. Sci.* **41:** 117-134.
- Schmitdtling, R. C., and Hiplins, V., 1998. Genetic diversity in longleaf pine (*Pinus palustris*) influence of historical and prehistorical events. Can. *Jour. For. Res.* 28: 1135–1145.
- Shang, H. Y., Wei, Y. M., Wang, X. R., and Zheng, Y. L., 2006. Genetic diversity and phylogenetic relationships in the rye genus *Secale* L. based on *Secale cereale* microsatellite markers. Genet. Mol. Biol. **29(4)**: 685-691.
- Sheidai, M., 2008. Comparative cytogenetic study of some grass genera of the subfamily Pooideae in Iran. *Polish Botanical Jour.* **53(1):** 15–28.
- Skizai, S., Rogalskal, S. M., and Boclanowski, J., 2007. RFLP analysis of mitochondrial DNA in the genus *Secale*. ACTA Biologica Cracoviensia Series Botanica, **49:** 77–87.
- Stoehr, M. U., and El-Kassaby, Y. A., 1997. Levels of genetic diversity at different stages of the domestication cycle of interior spruce in British Columbia. Theor. Appl. Genet. **94:** 83–90.
- Tamura, K., Dudley, J., Nei, M., and kumar, S., 2007. MEGA 4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. Mol. Biol. Evol. **24:** 1596-1599.
- Torkpo, S., Danquah, E., Offei, S., and Blay, E., 2006. Esterase, total protein and Seed storage protein diversity in Okra (*Abelmoschus esculentus* L.). West African J. Appl. Ecol. **9:** 7-18.
- Vaillancourt, A., Nkongolo, K. K., Michael, P., and Mehes, M., 2008. Identification, characterisation, and chromosome locations of rye and wheat specific ISSR and SCAR markers useful for breeding purposes. Euphytica, **159**: 297-306.
- Vences, F. J., Vaquero, F., and Perez de la Vega, M., 1987a. Phlogenetic relationships in *Secale*: An isozymatic study. Plant Syst. Evol. 157: 33-47.
- Vences, F. J., Vaquero, F., Garcia, P., and Perez de la Vega, M., 1987b. Further studies on phylogenetic relationships in *Secale*, On the origin of its species. Plant Breed. **98:** 281-291.
- Virk, D. S., Singh, D. N., Prasad, S. C., Gangwar, J. S., and Witcombe, J. R., 2003. Collaborative and consultative participatory plant breeding of rice for the rainfed uplands of eastern India. Euphytica. **132**: 95-108

- Vries, I. M., 1996. Characterization and identification of *Lactuca sativa* cultivars and wild relatives with SDS-electro phoresis (*Lactuca* sect. *Lactuca*, Compositae). Genet. Res. Crop Evol. **43:** 193–202.
- Wright, S., 1931. Evolution in Mendelian populations. Genetics, **16:** 97–159.
- Wright, S., 1951. The genetical structure of populations. Annals of Eugentics, **15**: 323–353.
- Wu, H. F., Li, Z. Z., and. Huang, H. W., 2006. Genetic differentiation among natural populations of *Gastrodia elata* (Orchidaceae) in Hubei and germplasm assessment of the cultivated populations. Biodiver. Sci. **14:** 315–326.
- Yeh, F. C., Yang, R. C., and Boyle, T., 1999. POPGENE version 1.32, Microsoft window base software for population genetic analysis: a quick user's guide. University of Alberta, Center for International Forestry Research, Alberta, Canada.
- Zhang, Q. Q., Xu, X. F., Qi, J., Wang, X. L., and Bao, Z. M., 2004. The genetic diversity of wild and farmed Japanese flounder populations. Periodical of Ocean University of China **34(5)**: 816–820.
- Zhou, Y. Q., Jing, J. Z., Li, Z. Y., Zhang, B. H., Wang, T. L., and Jia, J. F., 2005. ISSR identification of genetic diversity of *Rehmannia glutinosa* in Huai zone. Chinese Traditional Herbal Drugs, **36**: 257–262.
- Zou, C. Y., Li, L. J., Yang, K. C., Pan, G. T., and Ring, T. Z., 2010. Effects of Improvement by Mass Selection on the Different Maize Synthetic Populations. ACTA Agronomica Sinica, **36:** 76-84.

اثر انتخاب بر پارامترهای ژنتیکی Secale montanum بوسیله مارکر پروتئینهای ذخیرهای بذر ذخیرهای بذر

پروین صالحی شانجانی، استادیار، بانک ژن منابع طبیعی ایران، موسسه تحقیقات جنگلها و مراتع کشور، تهران، ایران (نویسنده مسئول) علی اشرف جعفری، استاد، بانک ژن منابع طبیعی ایران، موسسه تحقیقات جنگلها و مراتع کشور، تهران، ایران رویا حسین زاده، دانش آموخته کارشناسی ارشد، دانشگاه آزاد اسلامی واحد کرج، ایران

جكيده

خشک و نیمه خشک ارتفاعات ۲۹۰۰–۲۹۰۰ متری شمال و غرب ایران رویش دارد. در این مقاله الگو خشک و نیمه خشک ارتفاعات ۲۹۰۰–۲۹۰۰ متری شمال و غرب ایران رویش دارد. در این مقاله الگو پروتئینهای ذخیرهای بذر نه جمعیت وحشی S. montanum به همراه نتاج برتر و نمونه پلی کراس آنها مطالعه شدند. نتایج نشان دادند که جمعیتهای S. montanum دارای میانگین تعداد باند، هتروزیگوسیتی و پلیمورفیسم بالایی میباشند. گوناگونی باندهای نتاج برتر جمعیتهای مختلف کمتر از والدهای وحشی والدهای وحشیشان بود، درحالیکه پارامترهای ژنتیکی نمونه پلی کراس بسیار بیشتر از والدهای وحشی می یاشد. دو باند عمومی مختص به محل در الگو پروتئینهای ذخیرهای بذر تقریباً تمام جمعیتهای محتلف و نمونه پلی کراس وجود نداشت. این نتایج از فرضیه اثر انتخاب مصنوعی در کاهش تنوع ژنتیکی حمایت می کند. وجود اختلاف معنیدار در هتروزیگوسیتی مورد انتظار بین جمعیتهای وحشی و نتاج برترشان نشان می دهد که هرچه عملیات اصلاحی وسیعتر باشد فرسایش تنوع ژنتیکی نیز بیشتر می شود. دندروگرام حاصل از تجزیه کلاستر می گیرند. این نتایج نشان می دهد که اثر تعداد محدود والد و انتخابهای بعدی بیش از جدایی جغرافیایی می گیرند. این نتایج نشان می دهد که اثر تعداد محدود والد و انتخابهای بعدی بیش از جدایی جغرافیایی در تمایز ژنتیکی نقش دارد. نتایج این پژوهش نشان داد که مارکر پروتئینهای ذخیرهای بذر پتانسیل بالایی برای استفاده در برنامههای اصلاحی دارد.

کلمات کلیدی: Secale montanum، پروتئینهای ذخیرهای بذر، جمعیتهای وحشی، نتاج برتر